

# New Concepts in Pathogenesis of Renal Hypophosphatemic Syndromes

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During the past decades, our knowledge of renal phosphate handling has advanced dramatically. This advance is primarily due to the discovery of sodium-phosphate transport channels and their regulation in health and disease. The discovery of phosphatonins, initially in patients with tumor-induced osteomalacia, has not only allowed us to develop a better understanding of several rare diseases including vitamin D-resistant rickets, but also it has expanded our knowledge of the dynamic interaction between the bone and the kidney critical to bone mineralization. In this review, the author focuses on these new developments and their importance to our understanding of phosphate homeostasis in health and disease.

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## INTRODUCTION

Renal hypophosphatemic syndromes are a group of disorders characterized by phosphaturia due to inhibition of proximal tubular phosphate reabsorption. These disorders may present as isolated phosphaturia or as full-blown Fanconi syndrome. They could be due to a primary proximal tubular defect or presence of circulating factors with the ability to inhibit phosphate reabsorption by the proximal tubule. Finally, these syndromes could be hereditary or acquired in nature (Table). During the past decade significant progress has been made in our understanding of phosphate homeostasis in health and disease. This progress is partly due to advances in our understanding of rare acquired or hereditary diseases presenting with renal phosphate wasting. This review will focus primarily on these developments and their relevance to both normal phosphate homeostasis as well as renal hypophosphatemic disorders.

## BACKGROUND

Phosphorous, presenting primarily as inorganic phosphate, is the most abundant anion in human

body. It plays a major role in many biologic systems including cell membrane function, energy metabolism, cell signaling, and oxygen transport. The total body phosphorous pool is 99% intracellular, of which 85% is bound in the skeleton and the remainder in the soft tissues. The extracellular pool of phosphorous of approximately 600 mg exists primarily as  $\text{H}_2\text{PO}_4^-$  and  $\text{HPO}_4^{2-}$  with a ratio of 4 divalent to 1 monovalent molecule. The concentration of plasma inorganic phosphate, reported as the concentration of elemental phosphorous, is between 2.5 mg/dL to 4.5 mg/dL (range, 0.8 mmol/L to 1.4 mmol/L). Given the fact that the vast majority of phosphorous is intracellular, serum phosphorous is not a good indicator of the total body phosphate. In addition, the ratio of intracellular to extracellular phosphate is affected by multiple factors including acid-base status, glucose metabolism, and several hormones, especially insulin and catecholeamines.<sup>1,2</sup>

Phosphate intake, expressed as the amount of elemental phosphorous intake, is about 20 mg/kg/day or 1400 mg in a 70 kg man. Approximately, 80% of phosphorous intake (1100 mg) is absorbed

Classification of Renal Hypophosphatemic Syndromes\*

Cause	Disease
Primary proximal tubular disorder	
Hereditary	Mutation in phosphate transporters (Na-Pi-IIc transporter mutation [HHRH]) Fanconi syndrome due to inborn error of metabolism (Wilson disease and cystinosis)
Acquired	Drugs (tenefovir and isofosfamide) Toxins (lead and mercury) Tumor products (light-chain nephropathy and lysozymuria)
Presence of circulating factor/hormone	
Hereditary	X-linked hypophosphatemic rickets Autosomal dominant hypophosphatemic rickets Autosomal recessive hypophosphatemic rickets
Acquired	Primary hyperparathyroidism Tumor-induced osteomalacia Posthepatic resection hypophosphatemia

\*Na-Pi indicates sodium-phosphate and HHRH, hereditary hypophosphatemic rickets with hypercalciuria.

in the upper intestine, primarily in the proximal jejunum. This is counterbalanced by secretion of 200 mg of absorbed phosphorous in bile and saliva. The net absorption of phosphorous is therefore 900 mg or 64% of the total intake. In adults, but not in children, bone resorption and formation are equal and this dynamic process therefore has no impact on the overall phosphate homeostasis. In such individuals, renal excretion of phosphorous is equal to the net phosphorous absorption.

Intestinal phosphate absorption is both active (through sodium-phosphate transporters) and passive (by diffusion). Sodium-phosphate transporters, part of a large family of sodium-phosphate type II (Na-Pi-II) transporters, are located in the luminal membrane of both the renal proximal tubule and the upper intestine.<sup>3</sup> Intestinal phosphate absorption is regulated by the shuffling of Na-Pi-IIb transporters between the intracytoplasmic vesicles and luminal membrane. At low phosphate intake, most of the intestinal phosphate absorption occurs through active pathway, while at high phosphate intake, absorption mostly occurs passively.<sup>4</sup> Phosphate absorption is regulated by 1,25-vitamin D through the modulation of the number of sodium-phosphate transporters in the luminal membrane of enterocytes, primarily in the proximal jejunum.<sup>4</sup> As the activation of vitamin D is regulated by both parathyroid hormone (PTH) and serum phosphorous, phosphate absorption is therefore regulated through an interaction between serum phosphorous, PTH, and 1,25-vitamin D.<sup>5</sup>

## RENAL REGULATION OF PHOSPHATE

Phosphate is relatively freely filtered and then actively reabsorbed, resulting in a fractional excretion of about 10% to 15%. Phosphate reabsorption occurs primarily in the proximal tubule through Na-Pi-IIa, and to a lesser extent, Na-Pi-IIc transporters. By the end of the accessible proximal tubule, less than 30% of the filtered phosphate remains in the tubular fluid. The remainder of the filtered phosphate is reabsorbed in the distal portion of the proximal tubule with a small amount reabsorbed in the distal nephron. Phosphate is not secreted by any segment of the nephron.

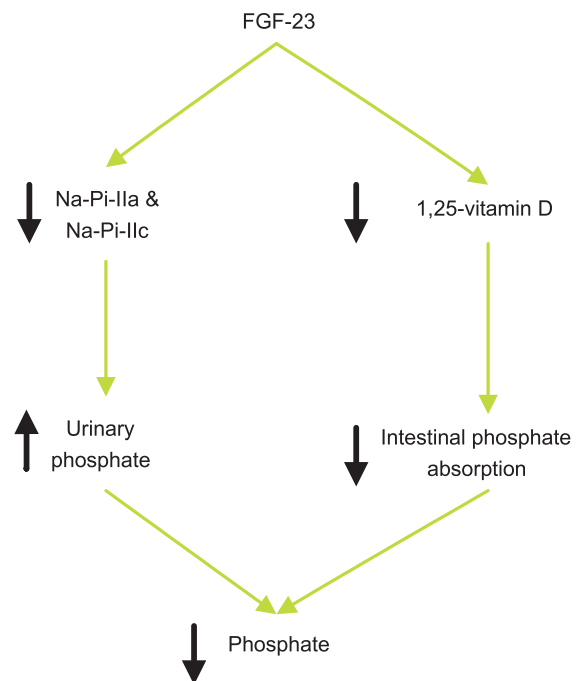
Major regulators of renal phosphate handling are PTH, dietary phosphate, and to a lesser extent, vitamin D. Parathyroid hormone, by binding to its receptor on the apical and basolateral membrane of the proximal tubules, decreases the abundance of Na-Pi-IIa and Na-Pi-IIc in the apical brush border membrane. Acutely, this is due to the movement of Na-Pi transporter from the brush border membrane to the intracellular pool. Long-term regulation is through the intracellular degradation of Na-Pi transporters. The basolateral membrane exit pathway for phosphate, however, remains unknown. Parathyroid hormone action is through activation of phospholipase C and protein kinase C, as well as protein kinase A pathways.<sup>6,7</sup> Dietary phosphate, independent of the PTH level, also rapidly modulates phosphate reabsorption in the proximal tubules through similar mechanisms.<sup>6,8</sup> The effect of vitamin D depends on its duration of administration as well as the state of the total body phosphate. Short-

term administration is phosphaturic in phosphate replete and antiphosphaturic in phosphate-depleted individuals.<sup>9,10</sup> Chronic administration of vitamin D increases gastrointestinal absorption of phosphate that secondarily increases urinary phosphate excretion.<sup>11</sup> Many other hormones also affect phosphate handling by the kidney. Growth hormone, insulin, and thyroid hormone increase, while calcitonin, glucocorticoids, and atrial natriuretic factor decrease phosphate reabsorption (reviewed in reference 6). These hormones and factors primarily exert their effect through regulation of Na-Pi-IIa transporter. Estrogen replacement is known to cause hypophosphatemia. This effect was very recently shown to be due to downregulation of Na-Pi-IIa transporter in the proximal tubules.<sup>12</sup>

### PHOSPHATONINS

In 1994, Cai and colleagues documented the presence of a circulating inhibitor of phosphate transport in the serum of a patient with tumor-induced osteomalacia (TIO). This patient presented with hypophosphatemia, phosphaturia, low 1,25-vitamin D, and rickets, which resolved with the removal of the tumor.<sup>13</sup> This circulating inhibitor was later shown to be fibroblast growth factor 23 (FGF-23).<sup>14</sup> The elevated serum level of this peptide is normalized with the removal of the tumor.<sup>15</sup> Fibroblast growth factor 23, a peptide with 251 amino acids, is synthesized by osteocytes. It is a member of a large family of FGFs that shows affinity for FGF receptors.<sup>16</sup> This peptide inhibits renal and intestinal absorption of phosphate directly by inhibition of Na-Pi-II transporters. It also inhibits activation of vitamin D to 1,25-vitamin D through inhibition of 1- $\alpha$ -hydroxylase enzyme directly (Figure 1).<sup>17,18</sup>

Patients with TIO have similar biochemical findings as 2 genetic syndromes, autosomal dominant hypophosphatemic rickets (ADHR) and X-linked hypophosphatemic rickets (XHR). In patients with ADHR, a mutated form of FGF-23 with prolonged half-life induces severe phosphaturia, hypophosphatemia, and rickets.<sup>19-21</sup> Patients with XHR have a mutation in a gene called *phosphate-regulating gene with homology to endopeptidases on X-chromosome (PHEX)*.<sup>22</sup> This gene is expressed primarily in the osteoblast lineage cells in the bone and the teeth.<sup>23</sup> Mutation



**Figure 1.** Fibroblast growth factor 23 (FGF-23) and regulation of serum phosphorous. Na-Pi IIa indicates sodium-phosphate type II a transporter.

in this gene is associated with an increase in serum concentration of FGF-23 in patients with XHR.<sup>18</sup> Although the mechanism by which *PHEX* regulates FGF-23 level remains controversial, the elevated level of FGF-23 in these patients is supportive of a mechanistic role for this factor in the development of hypophosphatemic rickets.<sup>18</sup> It is postulated that this enzyme normally breaks down FGF-23 and other phosphatonins, and therefore, an inactivating mutation could result in elevated levels of FGF-23. In a newly described hereditary disease, autosomal recessive hypophosphatemic rickets, mutation in another bone matrix protein, dentin matrix protein 1, is reported. These patients also have elevated levels of FGF-23 resulting in phosphaturia and rickets.<sup>24</sup> The mechanism by which this mutation leads to elevation in FGF-23 is unknown.

The physiological role of FGF-23 in normal individuals is still controversial. Dietary manipulation of phosphate intake results in either no change or only modest alterations in serum concentration of FGF-23.<sup>25,26</sup> An important regulator of FGF-23 is 1,25-vitamin D. In mice, administration of 1,25-vitamin D results in rapid upregulation of FGF-23 transcription in the

osteocytes, followed by an increase in its serum concentration. Osteoblast cells maintained in constant calcium and phosphate environment, increase their synthesis of FGF-23 in response to the administration of 1,25-vitamin D.<sup>27</sup> This effect creates a negative feedback loop which is important in the regulation of serum phosphorous. 1,25-vitamin D increases serum phosphorous by stimulating gastrointestinal absorption of phosphate, while at the same time, increasing concentration of FGF-23 results in inhibition of phosphate reabsorption by the kidney.<sup>25,26</sup> Hence, 1,25-vitamin D plays a dual and opposing role in phosphate homeostasis (Figure 2).

In patients with chronic kidney failure, serum levels of 1,25-vitamin D and FGF-23 are inversely correlated.<sup>28</sup> Fibroblast growth factor 23 in knocked out mice develop hyperphosphatemia and decreases renal excretion of phosphate.<sup>16</sup> These findings together support the concept that FGF-23 has an important physiologic role in phosphate homeostasis.

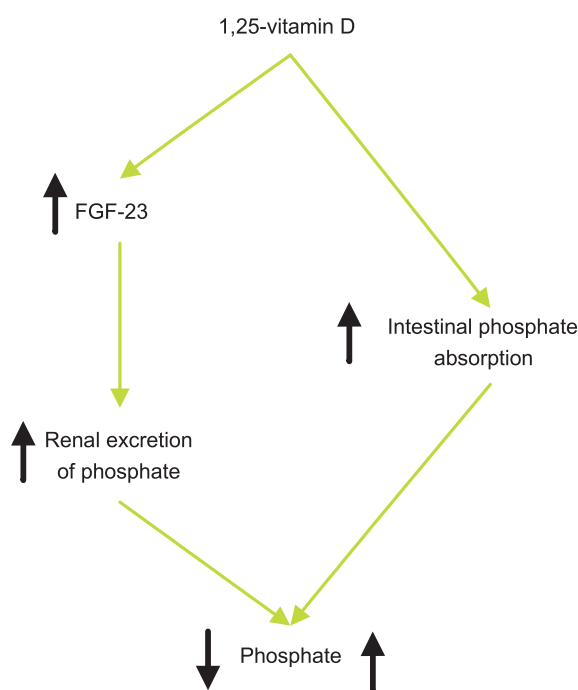
The mechanism of action of this peptide in the kidney is through interaction with FGF receptor on the basolateral membrane of the proximal tubule. This interaction requires presence of a unique

protein encoded by the *klotho* gene.<sup>29</sup> This interesting gene, known as anti-aging gene in mice, encodes 2 transcripts; a membrane-bound and a secreted protein.<sup>30</sup> Klotho protein acts as a cofactor for FGF-23 by binding to the FGF receptor in the basolateral membrane of the proximal tubule. This binding increases affinity of the receptor for FGF-23.<sup>29</sup> Mice deficient in klotho develop multiple defects including hyperphosphatemia due to a decrease in renal excretion of phosphate.<sup>31</sup> Recently, a patient has been reported with hypophosphatemic rickets and hyperparathyroidism due to a mutation resulting in elevated level of alpha-klotho protein.<sup>32</sup>

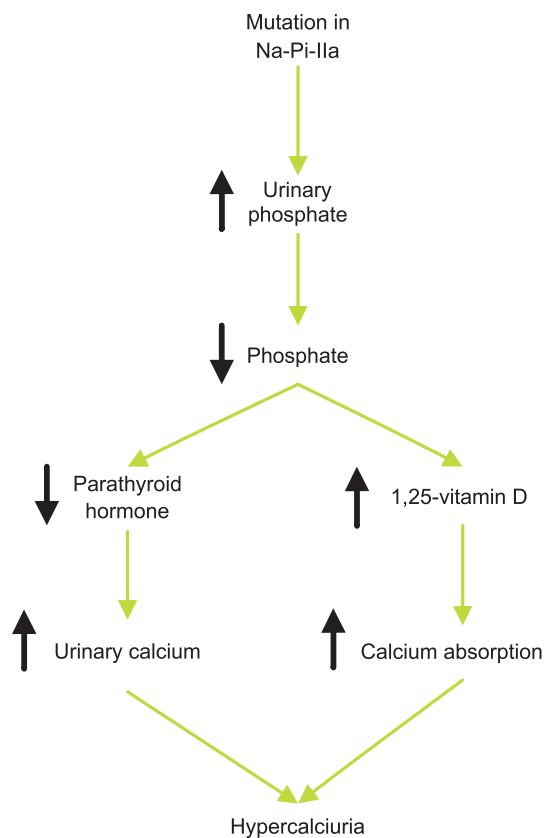
Several other phosphatonins, including matrix extracellular phosphoglycoprotein, frizzled-related protein 4, and FGF-7 are also secreted in TIO. These peptides are all phosphaturic through inhibition of Na-Pi transporters. The frizzled-related protein 4 inhibits the expected rise in 1,25-vitamin D level. In contrast, matrix extracellular phosphoglycoprotein infusion is associated with an increase in the level of this metabolite.<sup>17,18</sup> While these three peptides do contribute to hypophosphatemia seen in TIO as well as XHR, their role in phosphate homeostasis in the human is still speculative.

#### HYPOPHOSPHATEMIA DUE TO MUTATION IN SODIUM-PHOSPHATE TRANSPORTER

Hereditary hypophosphatemic rickets with hypercalciuria, first reported in Bedouin tribes, is a rare but interesting genetic disease that presents with renal hypophosphatemic rickets coupled with hypercalciuria. These patients, in contrast to those with XHR or ADHR, have appropriate increase in 1,25-vitamin D and decrease in PTH level. Recent work has shown a mutation in a candidate gene (*SLC34C3*) that encodes Na-Pi-IIc transporter. Although this transporter, which is located on the luminal membrane of renal proximal tubule, was thought to be responsible for a relatively small fraction of phosphate transport (about 15%), patients with homozygous or compound heterozygous mutation present with renal phosphate wasting. This finding therefore raises the possibility that Na-Pi-IIc transporter has a more important role in phosphate transport in human than previously thought. The resultant hypophosphatemia activates 1,25- $\alpha$ -hydroxylase enzyme resulting in increased level of 1,25-vitamin D. This would increase calcium absorption from the gastrointestinal tract inhibiting



**Figure 2.** Dual action of vitamin D on serum phosphate. FGF-23 indicates fibroblast growth factor 23.



**Figure 3.** Pathogenesis of hereditary hypophosphatemic rickets with hypercalciuria. Na-Pi IIa indicates sodium-phosphate type II a transporter.

PTH secretion and resulting in hypercalciuria (Figure 3).<sup>33,34</sup>

### SUMMARY

Our understanding of phosphate homeostasis has expanded rapidly by the discovery of a new class of phosphate-regulating hormones, the phosphatonins. These hormones, secreted by osteoblast-lineage cells, allow a dynamic dialogue between the bone and the kidney, through which they play a critical role in skeletal development. Such a discovery, while adding a newer layer to the complexity of calcium-phosphate homeostasis, has provided a molecular explanation for two enigmatic genetic diseases, ADHR and XHR. Further research is needed to better define the role of these hormones in phosphate homeostasis in health and disease, especially in patients with chronic kidney disease.

### CONFLICT OF INTEREST

None declared.

### REFERENCES

1. Gaasbeek A, Meinders AE. Hypophosphatemia: an update on its etiology and treatment. *Am J Med.* 2005;118:1094-101.
2. Amanzadeh J, Reilly RF, Jr. Hypophosphatemia: an evidence-based approach to its clinical consequences and management. *Nat Clin Pract Nephrol.* 2006;2:136-48.
3. Murer H, Forster I, Biber J. The sodium phosphate cotransporter family SLC34. *Pflugers Arch.* 2004;447:763-7.
4. Favus MJ. Intestinal absorption of calcium, magnesium and phosphorus. In: Coe FL, Favus MJ, editors. *Disorders of bone and mineral metabolism.* 2nd ed. New York: Lippincott Williams and Wilkins; 2002. p. 48-73.
5. Segawa H, Kaneko I, Yamanaka S, et al. Intestinal Na-P(i) cotransporter adaptation to dietary P(i) content in vitamin D receptor null mice. *Am J Physiol Renal Physiol.* 2004;287:F39-47.
6. Murer H, Hernando N, Forster I, Biber J. Proximal tubular phosphate reabsorption: molecular mechanisms. *Physiol Rev.* 2000;80:1373-409.
7. Bacic D, Lehir M, Biber J, Kaissling B, Murer H, Wagner CA. The renal Na<sup>+</sup>/phosphate cotransporter NaPi-IIa is internalized via the receptor-mediated endocytic route in response to parathyroid hormone. *Kidney Int.* 2006;69:495-503.
8. Keusch I, Traebert M, Löttscher M, Kaissling B, Murer H, Biber J. Parathyroid hormone and dietary phosphate provoke a lysosomal routing of the proximal tubular Na<sup>+</sup>/Pi-cotransporter type II. *Kidney Int.* 1998;54:1224-32.
9. Pastoriza-Munoz E, Mishler DR, Lechene C. Effect of phosphate deprivation on phosphate reabsorption in rat nephron: role of PTH. *Am J Physiol.* 1983;244:F140-9.
10. Stoll R, Kinne R, Murer H, Fleisch H, Bonjour JP. Phosphate transport by rat renal brush border membrane vesicles: influence of dietary phosphate, thyroparathyroidectomy, and 1,25-dihydroxyvitamin D<sub>3</sub>. *Pflugers Arch.* 1979;380:47-52.
11. Gloor HJ, Bonjour JP, Caverzasio J, Fleisch H. Resistance to the phosphaturic and calcemic actions of parathyroid hormone during phosphate depletion. Prevention by 1,25-dihydroxyvitamin D<sub>3</sub>. *J Clin Invest.* 1979;63:371-7.
12. Farouqi S, Levi M, Soleimani M, Amlal H. Estrogen downregulates the proximal tubule type IIa sodium phosphate cotransporter causing phosphate wasting and hypophosphatemia. *Kidney Int.* 2008;73:1141-50.
13. Cai Q, Hodgson SF, Kao PC, et al. Brief report: inhibition of renal phosphate transport by a tumor product in a patient with oncogenic osteomalacia. *N Engl J Med.* 1994;330:1645-9.
14. Shimada T, Mizutani S, Muto T, et al. Cloning and characterization of FGF23 as a causative factor of tumor-induced osteomalacia. *Proc Natl Acad Sci U S A.* 2001;98:6500-5.
15. Jonsson KB, Zahradnik R, Larsson T, et al. Fibroblast growth factor 23 in oncogenic osteomalacia and X-linked hypophosphatemia. *N Engl J Med.* 2003;348:1656-63.
16. Fukumoto S. Physiological regulation and disorders of phosphate metabolism—pivotal role of fibroblast growth

- factor 23. *Intern Med.* 2008;47:337-43.
17. Berndt TJ, Schiavi S, Kumar R. "Phosphatonins" and the regulation of phosphorus homeostasis. *Am J Physiol Renal Physiol.* 2005;289:F1170-82.
18. Berndt TJ, Kumar R. Phosphatonins and the regulation of phosphate homeostasis. *Ann Rev Physiol.* 2007;69:341-59.
19. AHDR Consortium. Autosomal dominant hypophosphataemic rickets is associated with mutations in FGF23. *Nat Genet.* 2000;26:345-8.
20. White KE, Carn G, Lorenz-Depiereux B, Benet-Pages A, Strom TM, Econs MJ. Autosomal-dominant hypophosphatemic rickets (ADHR) mutations stabilize FGF-23. *Kidney Int.* 2001;60:2079-86.
21. Bai XY, Miao D, Goltzman D, Karaplis AC. The autosomal dominant hypophosphatemic rickets R176Q mutation in fibroblast growth factor 23 resists proteolytic cleavage and enhances in vivo biological potency. *J Biol Chem.* 2003;278:9843-9.
22. [No authors listed]. A gene (PEX) with homologies to endopeptidases is mutated in patients with X-linked hypophosphatemic rickets. The HYP Consortium. *Nat Genet.* 1995;11:130-6.
23. Yuan B, Takaiwa M, Clemens TL, et al. Aberrant PheX function in osteoblasts and osteocytes alone underlies murine X-linked hypophosphatemia. *J Clin Invest.* 2008;118:722-34.
24. Lorenz-Depiereux B, Bastepe M, Benet-Pages A, et al. DMP1 mutations in autosomal recessive hypophosphatemia implicate a bone matrix protein in the regulation of phosphate homeostasis. *Nat Genet.* 2006;38:1248-50.
25. Ferrari SL, Bonjour JP, Rizzoli R. Fibroblast growth factor-23 relationship to dietary phosphate and renal phosphate handling in healthy young men. *J Clin Endocrinol Metab.* 2005;90:1519-24.
26. Sommer S, Berndt T, Craig T, Kumar R. The phosphatonins and the regulation of phosphate transport and vitamin D metabolism. *J Steroid Biochem Mol Biol.* 2007;103:497-503.
27. Liu S, Tang W, Zhou J, et al. Fibroblast growth factor 23 is a counter-regulatory phosphaturic hormone for vitamin D. *J Am Soc Nephrol.* 2006;17:1305-15.
28. Larsson T, Nisbeth U, Ljunggren O, Juppner H, Jonsson KB. Circulating concentration of FGF-23 increases as renal function declines in patients with chronic kidney disease, but does not change in response to variation in phosphate intake in healthy volunteers. *Kidney Int.* 2003;64:2272-9.
29. Kurosu H, Ogawa Y, Miyoshi M, et al. Regulation of fibroblast growth factor-23 signaling by klotho. *J Biol Chem.* 2006;281:6120-3.
30. Kuro-o M. Klotho as a regulator of oxidative stress and senescence. *Biol Chem.* 2008;389:233-41.
31. Segawa H, Yamanaka S, Ohno Y, et al. Correlation between hyperphosphatemia and type II Na-Pi cotransporter activity in klotho mice. *Am J Physiol Renal Physiol.* 2007;292:F769-79.
32. Brownstein CA, Adler F, Nelson-Williams C, et al. A translocation causing increased alpha-klotho level results in hypophosphatemic rickets and hyperparathyroidism. *Proc Natl Acad Sci U S A.* 2008;105:3455-60.
33. Bergwitz C, Roslin NM, Tieder M, et al. SLC34A3 mutations in patients with hereditary hypophosphatemic rickets with hypercalciuria predict a key role for the sodium-phosphate cotransporter NaPi-IIc in maintaining phosphate homeostasis. *Am J Hum Genet.* 2006;78:179-92.
34. Jaureguierry G, Carpenter TO, Forman S, Jueppner H, Bergwitz C. A novel missense mutation in SLC34A3 that causes HHRH identifies threonine 137 as an important determinant of sodium-phosphate co-transport in NaPi-IIc. *Am J Physiol Renal Physiol.* 2008;295:F371-9.

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