

# Association Between Visfatin Levels and Coronary Artery Disease in Patients with Chronic Kidney Disease

Yung-Chuan Lu,<sup>1</sup> Chia-Chang Hsu,<sup>2</sup> Teng-Hung Yu,<sup>3</sup>  
Chao-Ping Wang,<sup>3</sup> Li-Fen Lu,<sup>4</sup> Wei-Chin Hung,<sup>3</sup> Cheng-An Chiu,<sup>3</sup>  
Fu-Mei Chung,<sup>3</sup> Yau-Jiunn Lee,<sup>6</sup> I-Ting Tsai<sup>5</sup>

<sup>1</sup>Division of Endocrinology and Metabolism, E-Da Hospital, I-Shou University, Kaohsiung, Taiwan

<sup>2</sup>Division of Gastroenterology and Hepatology, E-Da Hospital, I-Shou University, Kaohsiung, Taiwan

<sup>3</sup>Division of Cardiology, Department of Internal Medicine, E-Da Hospital, I-Shou University, Kaohsiung, Taiwan

<sup>4</sup>Division of Cardiac Surgery, Department of Surgery, E-Da Hospital, I-Shou University, Kaohsiung, Taiwan

<sup>5</sup>Department of Emergency Medicine, E-Da Hospital, I-Shou University, Kaohsiung, Taiwan

<sup>6</sup>Lee's Endocrinologic Clinic, Pingtung, Taiwan

**Keywords.** visfatin, chronic kidney disease, coronary artery disease, atherosclerosis, E-selectin

**Introduction.** Visfatin (also known as pre-B cell colony-enhancing factor) is increased in patients with chronic kidney disease and has been linked with coronary atherosclerosis. Given that it has been reported that visfatin plays a role in endothelial dysfunction in chronic kidney disease patients, we examined associations between visfatin levels and several markers related to atherosclerosis.

**Materials and Methods.** The association between visfatin and atherosclerotic risk factors was studied in 173 chronic kidney disease patients (130 men and 43 women). Serum levels of visfatin were measured by the enzyme-linked immunosorbent assay.

**Results.** With increasing visfatin tertiles, patients proved to have a larger number of vessels with stenosis and a higher likelihood of coronary artery disease, as well as having incrementally lower estimated glomerular filtration rate and serum albumin and higher total leukocyte, neutrophil, and monocyte counts; high-sensitivity C-reactive protein; and brain natriuretic peptide levels. Visfatin showed significant positive correlations with low-density lipoprotein cholesterol, uric acid, blood urea nitrogen, creatinine, brain natriuretic peptide, E-selectin, total leukocyte count, neutrophil count, and high-sensitivity C-reactive protein, and a significant negative correlation with estimated glomerular filtration rate and albumin. Only E-selectin was independently associated with visfatin in multiple linear regression analysis.

**Conclusions.** This study indicates that plasma visfatin levels are significantly higher in the presence of coronary artery disease and are correlated with E-selectin levels, which suggest that increased plasma visfatin may be involved in the pathogenesis of coronary atherosclerosis in CKD patients.

IJKD 2013;7:446-52  
www.ijkd.org

## INTRODUCTION

Visfatin, also known as pre-B-cell colony-enhancing factor 1 (52 kDa to 55 kDa, middle molecule) or nicotinamide phosphoribosyltransferase (Nampt), is a ubiquitous adipokine that was first described by Fukuhara and colleagues<sup>1</sup> in 2005. Previous studies have suggested that visfatin may

be an important inflammatory protein and harmful factor in the setting of obesity-induced metabolic disturbance and associated with atherosclerosis, plaque destabilization in acute coronary syndrome, and carotid artery plaques.<sup>2-4</sup> Furthermore, our recent study showed that plasma visfatin levels were associated with infarct-related artery occlusion,

and also found a close association between visfatin and coronary artery disease (CAD).<sup>5</sup> The circulating levels of visfatin in patients with chronic kidney disease (CKD) have been reported to significantly increase and be correlated with endothelial dysfunction.<sup>6</sup> Thus, visfatin may be associated with the progression of atherosclerosis.

Systemic inflammation has been strongly implicated in both atherosclerosis and cardiovascular morbidity.<sup>7,8</sup> Patients with CKD are exposed to inflammation and endothelial dysfunction,<sup>9,10</sup> with inflammation playing a pivotal role in all stages of atherogenesis, from foam cell to plaque formation to rupture and ultimately to thrombosis. Insight gained from previous basic and clinical data linking inflammation to atherosclerosis has yielded important diagnostic and prognostic information.<sup>11,12</sup> Low-grade chronic inflammation as measured by high-sensitivity C-reactive protein (HS-CRP) predicts future risk of acute coronary syndrome independent of traditional cardiovascular risk factors.<sup>13</sup> Circulating levels of brain natriuretic peptide (BNP), a cardiac hormone, reflect the severity of cardiac dysfunction and inflammation.<sup>14</sup> An elevated total leukocyte count is a risk factor for atherosclerotic vascular disease, given that leukocyte-derived macrophages and other phagocytes are believed to contribute to vascular injury and atherosclerotic progression.<sup>15,16</sup> Increased expression of E-selectin is found in the endothelium of human atherosclerotic lesions.<sup>17</sup> However, an association of visfatin with atherosclerotic inflammatory markers and lipids in CKD patients has not yet been adequately investigated. In the present study, we examined the associations between visfatin levels and risk factors of coronary atherosclerosis.

## MATERIALS AND METHODS

### Patients

The study included 209 patients with mild-to-severe CKD (estimated glomerular filtration rate [GFR] between 89 mL/min/1.73 m<sup>2</sup> and 15 mL/min/1.73 m<sup>2</sup>) who underwent diagnostic coronary angiography for the examination of CAD due to typical and atypical chest pain at the Department of Cardiology of E-Da Hospital from June 2007 to December 2010. The GFRs were determined using the formula from the Modification of Diet in Renal Disease study immediately prior to the angiography

procedure. All patients with a GFR between 15 mL/min/1.73 m<sup>2</sup> and 89 mL/min/1.73 m<sup>2</sup> were eligible. Exclusion criteria were: (1) coronary artery bypass graft surgery history; (2) inflammatory diseases (such as infection, sepsis, malignancy, liver disease, and collagen disease); (3) steroid use or surgery within 1 month prior to admission; and (4) severe congestive heart failure (New York Heart Association classes III–IV). A total of 36 patients were excluded from the study: 13 patients had a positive history for coronary artery bypass graft surgery; 9 patients had inflammatory diseases; and 14 patients had class IV congestive heart failure. All 173 remaining patients were included in the study. This study was approved by the Human Research Ethics Committee of our hospital, and informed consent was obtained from each patient.

### Laboratory Measurements

Demographic data (age, sex, comorbidities, actual treatment, smoking status, body weight, and height) were collected before the angiographic procedure from the individual charts in the electronic hospital database. In the morning of the procedure day and after a 12-hour fasting period, blood samples were collected, stored, and analyzed by one laboratory. Serum levels of triglycerides, total cholesterol, low-density lipoprotein cholesterol (LDLC), high-density lipoprotein cholesterol (HDLC), uric acid, albumin, creatinine, and glucose were measured by standard commercial methods on a parallel, multichannel analyzer (Hitachi 7170A, Tokyo, Japan), as in our previous reports.<sup>18</sup> Peripheral leukocyte analyses included total leukocyte counts and differential percentages of neutrophils, monocytes, and lymphocytes using an automated cell counter (XE-2100 Hematology Alpha Transportation System, Sysmex Corporation, Kobe, Japan). The absolute count of a leukocyte subtype was calculated as the product of its respective differential percentage and total leukocyte count. To minimize the confounding effect of infection, subjects with a leukocyte count below  $4.0 \times 10^9/L$  or greater than  $10.0 \times 10^9/L$  were rechecked for analysis and examined extensively for possible occult of chronic infections. Any specimen with abnormal or atypical leukocytes were re-analyzed and excluded.

In addition, the concentrations of plasma visfatin and BNP were determined using a commercial enzyme immunoassay kit (Phoenix Pharmaceuticals,

Belmont, CA, USA). The concentrations of plasma C-reactive protein were measured using a high-sensitivity (HS-CRP) method (IMMAGE; Beckman Coulter, Immunochemistry Systems, Brea, CA, USA). E-selectin levels were determined by commercial solid phase enzyme linked immunosorbent assay kits (B-Bridge International, Sunnyvale, CA, USA, and Phoenix Pharmaceuticals, Belmont, CA and R & D Systems Inc, USA, respectively). The intra-assay coefficients of variation were 2.4% to 2.7% for visfatin, less than 5% for BNP, 4.2% to 8.7% for HS-CRP, and 5.2% to 6.6% for E-selectin. Samples were measured in duplicate in a single experiment.

### Angiographic Definitions

Coronary angiography images were obtained according to standard techniques, and the severity of stenosis was assessed using quantitative coronary angiography. Angiographies and quantitative coronary angiographic analysis were evaluated by at least 2 experienced interventional cardiologists blinded to clinical information and serologic parameters and were scored according to scoring system, the possible scores of this index ranged from 0 to 3 diseased vessels. The criterion for 1-, 2-, or 3-vessel disease was a greater-than-50% reduction in the internal diameter. The diameter of stenosis of the left main coronary artery could not exceed 50%.<sup>19</sup>

### Statistical Analysis

Data normality was analyzed using the Kolmogorov-Smirnov test. Continuous normally distributed variables are presented as mean  $\pm$  standard deviation, and non-normally distributed variables as median (interquartile range). Statistical differences in variables were compared using a 1-way analysis of variance for normally distributed variables followed by the Tukey pairwise comparison. Categorical variables were recorded as frequencies and percentages, and inter-group comparisons were analyzed by the chi-square test. Since the distributions of plasma visfatin, HS-CRP, E-selectin, BNP, and triglyceride were skewed, logarithmically transformed values were used for statistical analysis. The Pearson correlation analysis was used to examine the correlations between plasma visfatin and the values of other parameters. To assess the influence of tested parameters, multiple linear regression analysis was used. Statistical significance was accepted if the *P* value was less than .05. All of the statistical analyses were performed using SAS statistical software, version 8.2 (SAS Institute Inc, Cary, NC, USA).

## RESULTS

### Patients' Characteristics

Table 1 shows the clinical characteristics of

**Table 1.** Main Characteristics, According to Tertiles of Visfatin in 173 Patients With Chronic Kidney Disease\*

Parameter	Low Visfatin	Medium Visfatin	High Visfatin	<i>P</i>
Number of patients	57	58	58	...
Sex (male/female)				
Male	42	42	46	
Female	15	16	12	.66
Age, y	63.2 $\pm$ 12.9	63.3 $\pm$ 11.9	65.3 $\pm$ 13.9	.62
Hypertension	44 (77.2)	44 (75.9)	40 (69.0)	.56
Diabetes mellitus	30 (52.6)	20 (34.5)	19 (32.8)	.06
Hyperlipidemia	42 (73.7)	34 (58.6)	37 (63.8)	.23
Current smoking	28 (49.1)	26 (44.8)	31 (53.5)	.63
Body mass index, kg/m <sup>2</sup>	25.5 $\pm$ 4.4	25.3 $\pm$ 4.0	24.8 $\pm$ 3.8	.58
Waist-hip ratio	0.92 $\pm$ 0.06	0.94 $\pm$ 0.06	0.92 $\pm$ 0.05	.08
Systolic blood pressure, mm Hg	134 $\pm$ 26	132 $\pm$ 24	133 $\pm$ 27	.91
Diastolic blood pressure, mm Hg	76 $\pm$ 12	75 $\pm$ 10	76 $\pm$ 17	.96
Coronary artery disease	40 (70.2)	46 (79.3)	55 (94.8)	.003
Number of stenosed vessels				
0	17 (29.8)	12 (20.7)	3 (5.2)	
1	14 (24.6)	13 (22.4)	15 (25.9)	
2	11 (19.3)	15 (25.9)	19 (32.8)	
3	15 (26.3)	18 (31.0)	21 (36.2)	.048

\*Values in parentheses are percent.

173 CKD patients (male, 75.2%; female, 24.9%) stratified by visfatin. The mean visfatin level in the study was 28.5 ng/mL. The whole cohort median values of plasma visfatin levels were 13.4 ng/mL (interquartile range, 8.3 ng/mL to 26.0 ng/mL). The patients were divided according to tertiles of visfatin, as follows: low visfatin ( $\leq 10.2$  ng/mL),  $n = 57$ ; medium visfatin (10.2 ng/mL to 20.5 ng/mL),  $n = 58$ ; and high visfatin ( $\geq 20.5$  ng/mL),  $n = 58$ . Higher visfatin was associated with an increased prevalence of CAD and a larger number of stenosed vessels. Data presented in Table 2 show that with increasing visfatin tertiles, there was not only a significant decrease in albumin concentrations and GFR values, but also a significant increase in total leukocyte, neutrophil, and monocyte counts, HS-CRP, and BNP concentrations.

There were 71 patients with diabetic nephropathy, 48 with hypertensive nephropathy, 18 with chronic glomerular diseases, and 36 with unclassified causes of kidney failure. We also analyzed visfatin levels in patients stratified by these diseases. The mean plasma visfatin levels were  $32.4 \pm 63.8$  ng/mL,  $29.9 \pm 50.7$  ng/mL,  $23.1 \pm 20.8$  ng/mL, and  $25.0 \pm 24.9$  ng/mL in patients with diabetic nephropathy, hypertensive nephropathy, chronic glomerular diseases, and unclassified causes of kidney failure, respectively. There was no significant

difference in visfatin levels among the patients with different etiologies of CKD.

### Correlations Between Visfatin and Clinical And Biochemical Characteristics

The correlation analysis for circulating visfatin and relevant parameters are included in Table 3. Visfatin was positively correlated with LDLC, uric acid, blood urea nitrogen, creatinine, total leukocyte, neutrophil counts, HS-CRP, E-selectin, and BNP, and was negatively correlated with GFR and albumin. No association was found for body mass index, systolic and diastolic blood pressure, fasting blood glucose, hemoglobin A1c, total cholesterol, triglyceride, HDLC, hemoglobin, monocyte count, and lymphocyte count. In addition, multiple linear regression analysis was performed using visfatin as the dependent variable, and LDLC, uric acid, creatinine, albumin, total leukocyte and neutrophil counts, HS-CRP, E-selectin, and BNP as independent variables. Only E-selectin was independently associated with visfatin (Table 4).

### DISCUSSION

In the present study, we demonstrated in 173 CKD patients that visfatin showed significant positive correlations with LDL-cholesterol, uric acid, creatinine, BNP, E-selectin, total leukocyte

**Table 2.** Biochemical Characteristics, According to Tertiles of Visfatin in 173 Patients With Stages 2 to 4 of Chronic Kidney Disease\*

Parameter	Low Visfatin	Medium Visfatin	High Visfatin	P
Number of patients	57	58	58	...
Fasting blood glucose, mg/dL	152.4 $\pm$ 69.3	147.6 $\pm$ 82.9	145.5 $\pm$ 75.1	.90
Hemoglobin A1c, %	7.5 $\pm$ 1.9	6.8 $\pm$ 1.9	6.7 $\pm$ 1.9	.13
Total cholesterol, mg/dL	168.6 $\pm$ 45.4	173.0 $\pm$ 41.1	174.3 $\pm$ 40.4	.76
Triglyceride, mg/dL	119.5 (86.0 to 216.0)	109.0 (73.0 to 160.0)	97.0 (74.5 to 197.0)	.24
HDLC, mg/dL	36.6 $\pm$ 8.8	40.9 $\pm$ 9.9	39.0 $\pm$ 11.6	.09
LDLC, mg/dL	98.3 $\pm$ 35.7	107.9 $\pm$ 40.5	107.1 $\pm$ 36.6	.34
Uric acid, mg/dL	6.4 $\pm$ 1.8	6.5 $\pm$ 1.8	7.2 $\pm$ 2.6	.10
Blood urea nitrogen, mg/dL	18.7 $\pm$ 8.2	21.0 $\pm$ 11.4	23.5 $\pm$ 16.8	.14
Creatinine, mg/dL	1.3 $\pm$ 0.5	1.5 $\pm$ 1.0	1.6 $\pm$ 1.0	.09
GFR, mL/min/1.73 m <sup>2</sup>	62.3 $\pm$ 16.4	57.9 $\pm$ 19.2	54.2 $\pm$ 21.1	.047
Hemoglobin, g/dL	13.6 $\pm$ 1.8	12.7 $\pm$ 2.1	13.3 $\pm$ 2.3	.06
Albumin, g/L	4.0 $\pm$ 0.4	3.9 $\pm$ 0.4	3.8 $\pm$ 0.4	.03
Total leukocyte count, $\times 10^9/L$	7.848 $\pm$ 2.573	8.062 $\pm$ 3.017	10.240 $\pm$ 4.482	< .001
Neutrophil count, $\times 10^9/L$	4901 $\pm$ 2460	5173 $\pm$ 2408	8136 $\pm$ 4210	< .001
Monocyte count, $\times 10^9/L$	363 $\pm$ 130	461 $\pm$ 236	559 $\pm$ 415	.008
Lymphocyte count, $\times 10^9/L$	2137 $\pm$ 801	1983 $\pm$ 794	2025 $\pm$ 1340	.76
HS-CRP, mg/L	2.5 (1.2 to 8.4)	3.2 (1.0 to 7.3)	4.5 (1.4 to 25.1)	.04
E-selectin, ng/mL	24.6 $\pm$ 13.8	25.1 $\pm$ 11.9	27.5 $\pm$ 14.1	.58
BNP, ng/mL	4.6 (3.1 to 8.7)	6.4 (2.6 to 11.2)	5.4 (3.3 to 21.2)	.03

\*Data are expressed as mean  $\pm$  standard deviation or median (interquartile range). HDLC indicates high-density lipoprotein cholesterol; LDLC, low-density lipoprotein cholesterol; GFR, glomerular filtration rate; HS-CRP, high-sensitivity C-reactive protein; and BNP, brain natriuretic peptide.

**Table 3.** Correlation Between Plasma Visfatin and Relevant Parameters in 173 Patients With Chronic Kidney Disease

Variable	Visfatin Level	
	Pearson r	P
Body mass index	-0.035	.64
Systolic blood pressure	-0.036	.64
Diastolic blood pressure	-0.048	.53
Fasting blood glucose	0.017	.84
Hemoglobin A1c	-0.046	.60
Total cholesterol	0.071	.37
Triglyceride	-0.068	.38
High-density lipoprotein cholesterol	-0.023	.77
Low-density lipoprotein cholesterol	0.243	.03
Uric acid	0.195	.02
Blood urea nitrogen	0.205	.008
Creatinine	0.156	.04
Estimated glomerular filtration rate	-0.171	.02
Hemoglobin	-0.036	.64
Albumin	-0.209	.009
Total leukocyte count	0.245	.001
Neutrophil count	0.281	.001
Monocyte count	0.157	.08
Lymphocyte count	0.021	.811
High-sensitivity C-reactive protein	0.185	0.026
E-selectin	0.272	0.032
Brain natriuretic peptide	0.413	0.0001

**Table 4.** Multiple Linear Regression Analysis for Visfatin as a Dependent Variable

Variable	Estimate	Standard Error	P
Low-density lipoprotein cholesterol	-0.047	0.080	.57
Uric acid	0.608	1.673	.72
Creatinine	-12.240	12.583	.34
Albumin	-15.941	10.663	.15
Total leukocyte count	0.861	3.306	.80
Neutrophil count	0.000	0.004	.99
High-sensitivity C-reactive protein	-0.082	0.166	.63
E-selectin	0.874	0.354	.02
Brain natriuretic peptide	0.187	0.294	.53

count, neutrophil count, and HS-CRP, and a significant negative correlation with GFR and albumin. Further, a multiple regression analysis revealed that only E-selectin was independently associated with visfatin.

Water-soluble, protein-bound, and middle molecule uremic retention solutes were the three major groups of renal toxins. In a recent study, impaired renal filtration function was found to elevate circulating visfatin levels which belonged to the middle molecule uremic

retention solute and coronary atherosclerotic inflammatory marker.<sup>6,20</sup> The role of visfatin in nicotinamide adenine dinucleotide biosynthesis has been implicated in inflammatory states and this activity has been shown to be important for vascular smooth muscle cell maturation, indicating a possible involvement in vascular pathology.<sup>21,22</sup> In the present study, we have demonstrated a significant positive correlation between visfatin and total leukocyte count, neutrophil count, and HS-CRP, which concurs with the prior studies and suggest that there is a potential link between visfatin and inflammation.<sup>23,24</sup> Moreover, previous studies demonstrated that visfatin could activate human leukocyte expression of interleukin-1 $\beta$ , tumor necrosis factor- $\alpha$ , and especially interleukin-6,<sup>25</sup> as well as increase monocyte matrix metalloproteinase-9 activity in monocytic Tamm-Horsfall protein 1 cells.<sup>4</sup> Our previous study reported that plasma visfatin increased in patients with type 2 diabetes mellitus.<sup>26</sup> Filippatos and colleagues<sup>27</sup> also reported that plasma visfatin increased in patients with metabolic syndrome. Taken together, these important and intriguing results suggest that visfatin may be involved in the pathogenesis of atherosclerosis, diabetes and the metabolic syndrome, especially when these diseases are considered to be inflammatory processes.<sup>25,28</sup> It is therefore reasonable to propose that visfatin may act as a pro-inflammatory cytokine and that it plays a role in chronic inflammation, thus contributing to the pathogenesis of atherosclerosis and cardiovascular disease.<sup>3</sup>

E-selectin, one of the specific endothelial adhesion molecules playing an important role in the initiation of coronary atherosclerosis and acute coronary syndrome, has been well-investigated.<sup>29,30</sup> Endothelial dysfunction is one of the first hallmarks in the pathogenesis of atherosclerosis.<sup>31</sup> Endothelial injury may result in the release of various factors that can be detected in the circulation and can be potentially used as markers of endothelial dysfunction. Several studies have suggested that circulating adhesion molecules may serve as markers of endothelial damage or atherosclerosis. Noshad and coworkers proposed tissue endothelin-1 level as the main predicting factor of atherosclerosis.<sup>32</sup> The Atherosclerosis Risk In Communities study showed that E-selectin and intercellular adhesion molecule-1 are associated with carotid artery intima-

media thickness and are independent predictors of incident CAD.<sup>33</sup> Squadrito and colleagues<sup>34</sup> observed higher levels of circulating intercellular adhesion molecule-1 and E-selectin in patients with acute myocardial infarction. In the present study, E-selectin was significantly and positively correlated with visfatin, and was independently associated with visfatin in CKD patients by using multivariate analysis. Such a significant association between E-selectin and visfatin reiterates the findings of prior studies.<sup>35,36</sup> Furthermore, our current study found plasma visfatin levels to also significantly correlate with renal function parameters (blood urea nitrogen, serum creatinine, GFR, and serum albumin), metabolic risk factors (LDLC and uric acid) and BNP, which are strong independent and inverse predictors of cardiovascular events. This indicates and provides evidence that visfatin is synergistically increased with renal function deterioration and that both of these events contribute to coronary atherosclerosis.

## CONCLUSIONS

This study found that high plasma visfatin levels were significantly higher in the presence of CAD and correlated with E-selectin levels, which suggest that increased plasma visfatin may likely be involved in the pathogenesis of coronary atherosclerosis in CKD patients.

## ACKNOWLEDGMENTS

The authors would like to thank the E-Da Hospital of the Republic of China, Taiwan, for financially supporting this research under Contract EDAH100032. We would also like to thank the staff and members of the heart care team for their assistance in various measurements and other organizational aspects of this study.

## CONFLICT OF INTEREST

None declared.

## REFERENCES

1. Fukuhara A, Matsuda M, Nishizawa M, et al. Visfatin: a protein secreted by visceral fat that mimics the effects of insulin. *Science*. 2005;307:426-30.
2. Cheng KH, Chu CS, Lee KT, et al. Adipocytokines and proinflammatory mediators from abdominal and epicardial adipose tissue in patients with coronary artery disease. *Int J Obes (Lond)*. 2008;32:268-74.
3. Liu SW, Qiao SB, Yuan JS, Liu DQ. Association of plasma

visfatin levels with inflammation, atherosclerosis, and acute coronary syndromes in humans. *Clin Endocrinol (Oxf)*. 2009;71:202-7.

4. Dahl TB, Yndestad A, Skjelland M, et al. Increased expression of visfatin in macrophages of human unstable carotid and coronary atherosclerosis: possible role in inflammation and plaque destabilization. *Circulation*. 2007;115:972-80.
5. Yu TH, Lu LF, Hung WC, et al. Circulating visfatin level at admission is associated with occlusion of the Infarct-related artery in patients with acute ST-segment elevation myocardial infarction. *Acta Cardiologica Sinica*. 2011;27:77-85.
6. Malyszko J, Malyszko JS, Pawlak K, Mysliwiec M. Visfatin and apelin, new adipocytokines, and their relation to endothelial function in patients with chronic renal failure. *Adv Med Sci*. 2008;53:32-36.
7. Fischer-Betz R, Halle M, Schneider M. Inflammation-related cardiovascular morbidity: Pathophysiology and therapy. *Z Rheumatol*. 2010;69:680-4, 686-8.
8. Drakopoulou M, Toutouzas K, Michelongona A, Tousoulis D, Stefanadis C. Vulnerable plaque and inflammation: potential clinical strategies. *Curr Pharm Des*. 2011;17:4190-209.
9. Malyszko J. Mechanism of endothelial dysfunction in chronic kidney disease. *Clin Chim Acta*. 2010;411:1412-20.
10. Carmona AA, Rigon BG, Barroso MP, et al. Induction of systemic inflammation and thickening of subepicardial arteries in an animal model of uremia. *J Bras Nefrol*. 2011;33:408-12.
11. Shishehbor MH, Bhatt DL. Inflammation and atherosclerosis. *Curr Atheroscler Rep*. 2004;6:131-9.
12. Mohammadpour AH, Nazemian F, Moallem SA, Alamdaran SA, Asad-Abadi E, Shamsara J. Correlation between heat-shock protein 27 serum concentration and common carotid intima-media thickness in hemodialysis patients. *Iran J Kidney Dis*. 2011;5:260-6.
13. Ikeda U. Inflammation and coronary artery disease. *Curr Vasc Pharmacol*. 2003;1:65-70.
14. Quiroga B, Goicoechea M, García de Vinuesa S, et al. Cardiac markers in different degrees of chronic kidney disease: influence of inflammation and previous heart disease. *Med Clin (Barc)*. 2012;139:98-102.
15. Ernst E, Hammerschmidt DE, Bagge U, Matrai A, Dormandy JA. Leukocytes and the risk of ischemic diseases. *JAMA*. 1987;257:2318-24.
16. Fuster V, Lewis A. Mechanisms leading to myocardial infarction—insights from studies of vascular biology. *Circulation*. 1994;90:2126-46.
17. Davies MJ, Gordon JL, Gearing AJ, et al. The expression of the adhesion molecules ICAM-1, VCAM-1, PECAM, and E-selectin in human atherosclerosis. *J Pathol*. 1993;171:223-9.
18. Chung FM, Tsai JCR, Chang DM, Shin SJ, Lee YJ. Peripheral total and differential leukocyte count in diabetic nephropathy: The relationship of plasma leptin to leukocytosis. *Diabetes Care*. 2005;28:1710-7.
19. Wang CP, Hsu HL, Hung WC, et al. Increased epicardial

- adipose tissue (EAT) volume in type 2 diabetes mellitus and association with metabolic syndrome and severity of coronary atherosclerosis. *Clin Endocrinol.* 2009;70:876-82.
20. Axelsson J, Witasap A, Carrero JJ, et al. Circulating levels of visfatin/ Pre-B-Cell colony-enhancing factor 1 in relation to genotype GFR, body composition, and survival in patients with CKD. *Am J Kidney Dis.* 2007;49:237-44.
  21. van der Veer E, Ho C, O'Neil C, et al. Extension of human cell lifespan by nicotinamide phosphoribosyltransferase. *J Biol Chem.* 2007;282:10841-5.
  22. Archer SL. Pre-B-cell colony-enhancing factor regulates vascular smooth muscle maturation through a NAD<sup>+</sup>-dependent mechanism: recognition of a new mechanism for cell diversity and redox regulation of vascular tone and remodeling. *Circ Res.* 2005;97:4-7.
  23. Oki K, Yamane K, Kamei N, Nojima H, Kohno N. Circulating visfatin level is correlated with inflammation, but not with insulin resistance. *Clin Endocrinol (Oxf).* 2007;67:796-800.
  24. Kadoglou NP, Gkontopoulos A, Kapelouzou A, et al. Serum levels of vaspin and visfatin in patients with coronary artery disease-Kozani study. *Clin Chim Acta.* 2011;412:48-52.
  25. Moschen AR, Kaser A, Enrich B, et al. Visfatin, an adipocytokine with proinflammatory and immunomodulating properties. *J Immunol.* 2007;178:1748-58.
  26. Chen MP, Chung FM, Chang DM, et al. Elevated plasma level of visfatin/pre-B cell colony-enhancing factor in patients with type 2 diabetes mellitus. *J Clin Endocrinol Metab.* 2006;91:295-9.
  27. Filippatos TD, Derdemezis CS, Kiortsis DN, Tselepis AD, Elisaf MS. Increased plasma levels of visfatin/pre-B cell colonyenhancing factor in obese and overweight patients with metabolic syndrome. *J Endocrinol Invest.* 2007;30:323-6.
  28. Ross R. Atherosclerosis—an inflammatory disease. *New Engl J Med.* 1999;340:115-26.
  29. Dong ZM, Chapman SM, Brown AA, Frenette PS, Hynes RO, Wagner DD. The combined role of P- and E-selectins in atherosclerosis. *J Clin Invest.* 1998; 102:145-52.
  30. Suefuji H, Ogawa H, Yasue H, et al. Increased plasma level of soluble E-selectin in acute myocardial infarction. *Am Heart J.* 2000;140:243-8.
  31. Vanhoutte PM. Endothelial dysfunction and atherosclerosis. *Eur Heart J.* 1997;18:E19-23.
  32. Noshad H, Argani H, Nezami N, et al. Arterial atherosclerosis in patients with chronic kidney disease and its relationship with serum and tissue endothelin-1. *Iran J Kidney Dis.* 2009;3:203-9.
  33. Salomaa V, Stinson V, Kark JD, Folsom AR, Davis CE, Wu KK. Association of fibrinolytic parameters with early atherosclerosis. The ARIC Study. Atherosclerosis Risk in Communities Study. *Circulation.* 1995;91:284.
  34. Squadrito F, Saitta A, Altavilla D, et al. Thrombotytic therapy with urokinase reduces increased circulating endothelial adhesion molecules in acute myocardial infarction. *Inflamm Res.* 1996;45:14-9.
  35. Malyszko J, Malyszko JS, Pawlak K, Mysliwiec M. Visfatin and apelin, new adipocytokines, and their relation to endothelial function in patients with chronic renal failure. *Adv Med Sci.* 2008;53:32-6.
  36. Lee WJ, Wu CS, Lin H, et al. Visfatin-induced expression of inflammatory mediators in human endothelial cells through the NF-kappaB pathway. *Int J Obes (Lond).* 2009;33:465-72.

Correspondence to:  
 I-Ting Tsai, MD  
 Department of Emergency, E-Da Hospital, I-Shou University,  
 No. 1, Yi-Da Rd, Jiau-Shu Village, Yan-Chao Township,  
 Kaohsiung, 82445, Taiwan  
 Tel: +886 7 615 1100 ext. 5914 or 5018  
 Fax: +886 7 375 7678  
 E-mail: gene6623@yahoo.com.tw

Received October 2012  
 Revised April 2013  
 Accepted May 2013